

Use of suspensions of phytosterol microparticles to improve the solubility of methane in water[☆]

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Abstract – Methane, an abundantly produced greenhouse gas, is a major driver of global climate change. The development of sustainable processes for methane capture is limited by the poor solubility of this organic compound in water, but this solubility can be increased by adding other compounds to the aqueous solution. Here, we studied the solubility of methane in aqueous dispersions containing lipid microparticles of phytosterols—a group of sterol compounds from plants—at room temperature and atmospheric pressure. The solubility of the phytosterols in water was improved chemically, by functionalization with glycerol carbonate in solvent-free conditions, and physically, by antisolvent precipitation to form particles in water. We evaluated the influence of phytosterol microparticles on methane solubility in water by monitoring the apparent partition coefficient between the gas and liquid phases defined by Henry’s law. Phytosterol functionalization generated amphiphilic phytosterols with glycerol branches, which were characterized by FTIR and MALDI-TOF. These amphiphilic phytosterols formed particles of 2 and 12 μm in diameter on antisolvent precipitation. Systems containing phytosterol microparticles had a partition coefficient half that of the corresponding control. The gas-liquid equilibrium was, therefore, shifted to the liquid state, demonstrating that the solubility of methane in water was improved by phytosterol microparticles.

Keywords: Phytosterols / methane solubility / functionalization / Henry’s constants

Résumé – **Utilisation de suspensions de microparticules de phytostérols pour améliorer la solubilité du méthane dans l’eau.** Le méthane, un gaz à effet de serre produit en abondance, est un facteur majeur du changement climatique mondial. Le développement de procédés durables de captage du méthane est limité par la faible solubilité de ce composé organique dans l’eau, mais cette solubilité peut être augmentée en ajoutant d’autres composés à la solution aqueuse. Ici, nous avons étudié la solubilité du méthane dans des dispersions aqueuses contenant des microparticules lipidiques de phytostérols – un groupe de composés stérols issus de plantes – à température ambiante et à pression atmosphérique. La solubilité des phytostérols dans l’eau a été améliorée chimiquement, par fonctionnalisation avec du carbonate de glycérol dans des conditions sans solvant, et physiquement, par précipitation pour former des particules dans l’eau. Nous avons évalué l’influence des microparticules de phytostérols sur la solubilité du méthane dans l’eau par détermination de son coefficient de partage apparent entre les phases gazeuse et liquide comme défini par la loi de Henry. La fonctionnalisation des phytostérols a généré des phytostérols amphiphiles, caractérisés par FTIR et MALDI-TOF. Ces phytostérols amphiphiles forment dans l’eau des particules de 2 et 12 μm de diamètre par précipitation dans un anti-solvant. Les systèmes contenant des microparticules de phytostérols présentent un coefficient de partage moitié inférieur à celui du contrôle. L’équilibre gaz-liquide a donc été déplacé vers l’état liquide, démontrant que la solubilité du méthane dans l’eau est améliorée par les microparticules de phytostérols.

Mots clés : Phytostérols / solubilité du méthane / fonctionnalisation / constantes de Henry

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Highlights

- Microparticles of phytosterol improve the solubility of methane in aqueous solution.
- Amphiphilic phytosterols were synthesized from glycerol carbonate and phytosterols.
- Methane showed a better affinity with systems containing amphiphilic phytosterols than native phytosterols.
- Pure olive oil had the best capacity for CH₄ absorption, with the lowest gas-liquid partition coefficient H.

1 Introduction

Methane (CH₄) is a powerful molecule with several applications in energy production, heat production, and as a biofuel (Chen and Weng, 2023; Freakley *et al.*, 2021). However, it is also a major driver of global climate change as one of the most abundantly produced greenhouse gases (GHG), accounting for 30% of current net climate forcing (Alonso *et al.*, 2017). It has a global warming potential 28 times greater than that of carbon dioxide (CO₂) over a 100-year reference time frame (Freakley *et al.*, 2021). The technological development of CH₄ recovery processes is a major issue in the chemical industry and in biogas production. Research is currently focusing on new processes for minimizing atmospheric CH₄ emissions (Lackner, 2020). Current solutions for the physical removal of CH₄ include cryogenic separation, molecular sieves and zeolite filters (Boucher and Folberth, 2010), all of which are very costly in terms of energy and technology. However, methane capture methods are limited by the low solubility of methane in water (Sander, 2008). There are two ways of improving this solubility. The first involves reacting methane with other substances to form products that dissolve more readily in water (Horn and Schlögl, 2015). The second involves dissolving methane in solution with a suitable reagent (Avalos Ramirez *et al.*, 2012). Rigorous technical and economic data are also currently lacking for CH₄ capture with these methods.

The chemical conversion of methane requires high temperatures and pressures due to its strong C–H bond (434 kJ/mol) (Cai and Hu, 2019). Under catalytic action, methane can be converted into water-soluble chemicals, such as methanol (Hameed *et al.*, 2014), carboxylic acid (Periana *et al.*, 2003), and aromatic hydrocarbons (Tang *et al.*, 2014) under specific reaction conditions. The high energy barrier required for methane activation necessitates the use of extreme reaction conditions—with high temperatures, high pressures, and strong acids—for the conversion of methane into other chemicals (Skutil and Taniewski, 2006).

It is also possible to use methanotrophs—microorganisms that use methane as a carbon source—to transform CH₄ into molecules of interest. However, the use of such microorganisms in CH₄ capture is limited by the poor solubility of CH₄ in aqueous medium. Several studies have reported the use of oily components in the medium to improve yields. For example, the addition of paraffin oil to water has been shown to

increase methane solubility 10-fold (Han *et al.*, 2009), with a 1.6 times higher methanol yield from methanotrophs due to the increase in methane availability (Patel *et al.*, 2020).

The identification of a suitable solution that can dissolve methane without transformation reactions is challenging. Several methods have been developed for increasing the solubility of poorly soluble organic compounds such as methane, by adding compounds, such as surfactants, to the medium. Methane solubility decreases with increasing temperature and increases linearly with the concentration of sodium dodecyl sulfate (SDS) above the critical micelle concentration (CMC) or critical aggregation concentration (CAC) of the surfactant (Valentin and Mouloungui, 2013), indicating that the gas molecules are solubilized by the micelles in these solutions (Hai *et al.*, 1999). Standard Gibbs energies for the transfer of methane from bulk solutions to micelles have large negative values, indicating that the hydrophobic gas is present preferentially in the hydrophobic interior of the micelles (Hariz *et al.*, 2017). Micelles of sodium oleate and cyclohexane at a molar ratio of 1/3 form a nanometric system (optimum for micelles of 200–350 nm in diameter) for the solubilization of methane at concentrations of up to 17.1 ml CH₄/mL water (Huang *et al.*, 2019). Another study by the same team (Huang *et al.*, 2020) showed that surfactant-alkane systems, such as that based on sodium oleate/hexane, had remarkable solubility properties, superior to those of systems based solely on surfactants.

One study showed that the optimal conditions for methane solubilization in water to which anionic–nonionic surfactants were added at various ratios was variable (Zhou *et al.*, 2018). Based on an analysis of the solubilization effect and the stability of different absorbents, it was concluded that anionic–nonionic surfactant system were more effective for methane solubilization than other selected surfactants. King (1992) measured methane solubility in aqueous solutions of a commercially available polyethoxylated lauryl alcohol, for aqueous solutions containing two different mixtures of BRIJ 35 and BRIJ 30. The intracellular solubility of methane gas was found to increase with surfactant concentration, for both pure BRIJ 35 and for each of the surfactant mixtures. In terms of methane solubilization, it was concluded that polar polymerized ethylene oxide groups do not contribute to the solubilization of methane and that the solubilizing capacities of the hydrophobic interiors of the micelles composed of BRIJ 35 and its mixtures with BRIJ 30 closely approximate the sorptive capacity of pure lauryl alcohol. The solubility of methane in solutions of monoethanolamine, diethanolamine, and triethanolamine has been measured at temperatures of 25–125°C and pressures of up to about 13 MPa, and methane was found to be soluble (molar fraction) in the amine solution than in pure water. Furthermore, solubility increased with the size of the alkanolamine (Carroll *et al.*, 1998). The use of nanoparticles as sorbent materials for methane capture has also attracted considerable interest. These nanomaterials have a high surface-to-volume ratio for interactions and can be synthesized to have specific physicochemical properties. Several studies have suggested that organic nanomaterials, such as phenol-based activated carbon (Lee *et al.*, 2007), carbon MW-nanotubes preloaded with water (Zhou *et al.*, 2005), or metal organic frameworks (MOF), could be used for CH₄ capture (Llewellyn *et al.*, 2008; Saha *et al.*, 2010). It is difficult to

compare studies due to differences in the pressures and temperatures used, but the studies cited above suggest that these nanomaterials increase the solubility of methane in water.

We focus here on phytosterols because they are biosourced and biodegradable compounds biologically synthesized from squalene. Squalene has been shown to have a good affinity for methane (Chappelow and Prausnitz, 1974), but to the best of our knowledge, phytosterols affinity with methane has never reported in the literature. Overall, there are no studies demonstrating the increase in the solubility of methane in water under the action of phytosterols and derivatives. Squalene is an hydrophobic molecule able to solubilize other hydrophobic organic compounds. Phytosterols are triterpenes with a hydrophobic skeleton consisting of a four-membered steroid ring with a side chain of 9 or 10 carbon atoms. Their hydrophilic moiety results from the presence of an OH group, providing possibilities for chemical modification of potential value for industrial development (Mouloungui *et al.*, 2006). Phytosterols could be a cheaper alternative to squalene to solubilize methane. Most of the phytosterols currently on the market are obtained as by-products of the production and refining of oils from the seeds of plants such as soybean and rapeseed. Phytosterols are obtained from the recycling of deodorization wastewater (Daguet and Coic, 1999). Like squalene and squalene, phytosterols are insoluble in water and require specific preparation to form stable dispersions in water. They can form nanoparticles stabilized by a surfactant by the antisolvent precipitation method (Rossi *et al.*, 2010).

Structural modifications to phytosterols are primarily designed to increase solubility in oils, but phytosterols also have considerable potential for applications in low-fat and fat-free foods if their water solubility is increased. Physical modifications (mainly micro-encapsulation) can be used to render phytosterols more hydrophilic. Alternatively, chemical modifications in which polar components are added to phytosterols can be used to obtain chemically stable phytosterols that are more soluble in water. There have been far fewer studies of synthetic hydrophilic phytosterol derivatives than of lipophilic phytosterol derivatives (*e.g.* phytosterol esters, etc.). A recent review (Hu *et al.*, 2022) summarized recent progress in the field of hydrophilic phytosterol derivatives. Most hydrophilic modifications of phytosterols involve the introduction of polar components at the C-3 hydroxyl group. There are two main types of strategy for synthesizing hydrophilic phytosterol derivatives: (1) direct linking of polar components, as in ethoxylated phytosterols (Folmer, 2003), phytosterol glycosides and phytosterol amino acid esters; and (2) coupling with polar components via a spacer, as in phytosterol polyethylene glycol succinate and phytosterol sorbitol adipate. Spacers may be flexible and can change the properties of the entire molecule. Polybasic organic acid and binary anhydride spacers are the most widely used.

In this study, we investigated the influence of phytosterols on CH₄ solubility in water in order to use them as new, biosourced and biodegradable additives for CH₄ solubilization. Two phytosterol systems were tested: direct dispersion in water with raw mixing (RM), and by antisolvent precipitation (ASP). In both cases, two types of phytosterols were used: native and chemically modified phytosterols. The antisolvent method, as described by Rossi *et al.* (2010), generated phytosterol microparticles, and chemically modified

phytosterols were obtained with glycerol carbonate—a biosourced glycerol derivative —, according to a method based on previously reported protocols (Holmiere *et al.*, 2017; Valentin and Mouloungui, 2013). We also studied the influence of olive oil and epoxidized soy oil on CH₄ solubilization as both have been reported to contain 288 mg/100 g and 355 mg/100 g of phytosterols respectively (Yang *et al.*, 2019). We used measurements of Henry's law constant H to follow the changes in methane solubility in the different aqueous phases using an innovative experimental setup (Benizri *et al.*, 2017). We sought to reduce the value of the H(CH₄) constant and to improve selectivity for methane relative to carbon dioxide.

2 Experimental procedures

2.1 Materials

Glycerol carbonate (CAS 931-40-8, Hunstman), ethanol (HPLC grade, CAS 64-17-5, VWR), ethyl acetate (HPLC grade, CAS 141-78-6, VWR), cyclohexane (HPLC grade, CAS 110-82-7, VWR) and ZnSO₄·H₂O (CAS 7446-19-7, Sigma Aldrich) were purchased and used directly. Phytosterols were obtained from BASF (CAS 949109-75-5, GENEROL 98 RF BASF) and their composition was determined by gas chromatography (β-sitosterol (42.2%), campesterol (33.5%), brassicasterol (8.1%), stigmasterol (0.4%)). Epoxidised soy oil (V-ZICLUS 65 “ESBO”, lot 22BT0504A1, iodine value of 1.6 g(I₂)/100 g, and oxirane value of 6.6 g(O₂)/100 g) was obtained from Quimidroga (Spain). Commercial olive oil was obtained from Carrefour (Carrefour Extra).

2.2 Synthesis of amphiphilic phytosterols

Amphiphilic phytosterols (PS-amph) were synthesized in a 1 L-batch reactor with mechanical stirring, a refrigerant and a thermometer. The reactor was heated at 220 °C and stirred at 500 rpm. Phytosterols (27 g) and glycerol carbonate (GC, 740 g) were added to the reactor. Once the phytosterols had completely melted, 10 g ZnSO₄ was added as a catalyst. The reaction lasted at least 7 h. PS-amph were collected by liquid-liquid extraction with ethyl acetate as the organic solvent, and water as the aqueous solvent to eliminate the ZnSO₄, glycerol and glycerol carbonate. Both organic and aqueous phases were treated twice, and the ethyl acetate from the organic phase was evaporated off for collection of the PS-amph in solid form.

2.3 Characterization of amphiphilic phytosterols

PS-amph were analyzed by mass spectrometry in chloroform by matrix-assisted laser desorption ionization (MALDI-TOF). Spectra were recorded in reflection mode on a DSQII (Thermo Fisher Scientific) for analyses in chemical desorption-ionization NH₃ mode at Université Paul Sabatier (Toulouse, France). Fourier transform infrared (FTIR) spectra of PS-amph were obtained with a Perkin Elmer Spectrum 65 spectrometer equipped with an attenuated total reflectance (ATR) device (Specac) with a single-reflection diamond crystal.

2.4 Production of phytosterol dispersions

2.4.1 Raw mixing (RM)

Native phytosterols (**RM-native**) or PS-amph (**RM-amph**) were dispersed in demineralized water at 16 g/L by vigorous stirring with a Silverson L4RT at 7500 rpm for 3 min.

2.4.2 Antisolvent precipitation (ASP)

Samples **ASP-native 1** and **ASP-native 5** were prepared at 1 g/L and 5 g/L respectively by the ASP method as described by Rossi *et al.* (2010). Phytosterols were first dissolved in an organic solvent to form the organic phase. This phase was then mixed with another solvent, miscible with the organic solvent, but in which phytosterols were not soluble, causing precipitation. For **ASP-native 1**, we dissolved 2 g phytosterol in 200 g absolute ethanol plus 5.78 g Tween 80. After complete solubilization, this organic phase was rapidly added to 1800 g demineralized (ratio org:aq of 1:9V/V) water with vigorous stirring with a Silverson L4RT at 20,000 rpm. For **ASP-native 5**, 10 g phytosterol were dissolved in 1000 g absolute ethanol plus 28.9 g Tween 80. After complete solubilization, this organic phase was rapidly added to 1000 g demineralized water (ratio org:aq of 1:1V/V) with stirring at 20,000 rpm.

Samples **ASP-amph 1**, **ASP-amph 5** and **ASP-amph 15** were prepared according to the following procedure. For dispersions at 1 g/L (**ASP-amph 1**), 1 g PS-amph was dissolved in 100 g absolute ethanol. After complete solubilization, 40 g of this organic phase was rapidly added to 360 g demineralized water (ratio org:aq of 1:9 V/V) with vigorous stirring with a Silverson L4RT at 7500 rpm. Stirring was continued for another three minutes. For dispersions at 5 g/L (**ASP-amph 5**) and 15 g/L (**ASP-amph 15**), 2 g and 6 g PS-amph, respectively, was dissolved in 200 g absolute ethanol. After complete solubilization, the organic phase was rapidly added to 200 g demineralized water (ratio org:aq of 1:1 V/V) with vigorous stirring with a Silverson L4RT at 7500 rpm. Stirring was continued for another 3 min.

Both types of dispersions were analyzed by dynamic light scattering (DLS) with a Malvern MS 3000 to evaluate particle size.

2.5 Measuring Henry constants with a saturometer

2.5.1 Procedure for measuring Henry's constants

At low pressures, the solubility of gases in liquids is governed by Henry's law, according to which the saturated solubility of a gas in a liquid is proportional to its partial pressure. The affinity of methane for different solutions was therefore evaluated by calculating the Henry's law constant H . Lower values of H indicate a greater absorption in the liquid phase. This constant was measured with the "saturometer" experimental setup (Benizri *et al.*, 2017) developed and marketed by EPURTEK (Lacroix-Falgarde, FRANCE). This setup is a batch contactor for gas and liquid in which temperature, pressure and phase concentration are measured continuously or sequentially. The composition of the gas phase (CH_4 and CO_2) was determined by gas chromatography with a thermal conductivity detector.

The procedure followed was that described in Benizri *et al.* (2017). Briefly, the tank for the liquid phase was filled

with the appropriate liquid (water, or phytosterol dispersions) at atmospheric pressure. The gas tank was then filled with CH_4 or CO_2 and purged for 3 min. The pressure in the gas cell was initially higher than atmospheric pressure and was adjusted before taking a sample and recording the temperature. The two tanks were separated by a valve, which was closed before the gas injection. This valve was then opened to allow contact between gas and liquid, and the whole system was stirred by rotation of the apparatus until equilibrium was reached. Once the pressure had stabilized, the valve was closed and the temperatures of the gas and liquid were recorded before taking new samples. All measurements occurred at temperatures between 20 and 25 °C. Experiments were performed three times to ensure repeatability.

2.5.2 Analysis of gas composition

Gas samples were analyzed by gas chromatography with a Trace 1300 gas chromatograph equipped with a thermoconductivity detector (TCD). Samples were injected into a PLOT capillary column with a PTFE filter at the injection valve to prevent contamination of the column with liquid and impurities. Helium was used as the carrier gas and the flow rate for the injection was 1 mL/min in split-less mode.

2.5.3 Henry's constant calculation

The initial and equilibrium molar fractions of the compounds in the liquid and gas phases can be determined from a material balance. The quantity of matter transferred from the gas to the liquid is denoted $n_{i,trans}$ and is calculated as follows (Eq. (1)):

$$n_{i,trans} = n_{gas,initial} - n_{gas,final} = \frac{y_{i,in} \times P_{s,in} \times V_{gas}}{R \times T_{gas,in}} - \frac{y_{i,eq} \times P_{s,eq} \times V_{gas}}{R \times T_{gas,eq}} \quad (1)$$

where

- $y_{i,in}$ is the molar fraction of compound i in the gas phase in the initial state
- $y_{i,eq}$ is the molar fraction of compound i in the gas phase at equilibrium
- $P_{s,in}$ is the pressure of the gas in its initial state
- $P_{s,eq}$ is the pressure of the gas at equilibrium
- V_{gas} is the volume of the gas cell
- R is the perfect gas constant
- $T_{gas,in}$ is the temperature of the gas in its initial state
- $T_{gas,eq}$ is the temperature of the gas at equilibrium

In this equation, the dissolved gas input is assumed to be negligible relative to the number of moles of water. The pressure and temperature values are recorded by the sensor before and after gas–liquid contact. The gas fraction of compound i is determined by gas chromatography, and the molar fraction in the liquid is defined by the relationship (Eq. (2)):

$$x_i = \frac{n_{i,trans}}{n_{liq}} = \frac{n_{i,trans} \times M_{liq}}{\rho_{liq} V_{liq}} \quad (2)$$

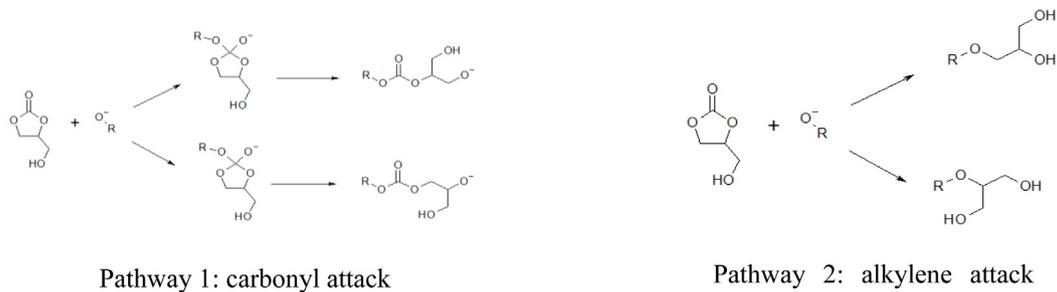


Fig. 1. Reaction mechanisms for 5-membered carbonate ring opening (Ochoa-Gómez *et al.*, 2012).

By Henry's law, the apparent partition coefficient H of a compound i is calculated as follows (Eq. (3)):

$$H_i = \frac{y_i \times P_{eq}}{x_i} = \frac{y_{i,eq} \times P_{s,eq} \times n_{liq}}{n_{i,trans}} = \frac{y_{i,eq} \times P_{s,eq} \times \rho_{liq} \times V_{liq}}{n_{i,trans} \times M_{liq}} \quad (3)$$

As the apparent Henry's law constant of a compound decreases, its absorption by the liquid phase increases and it becomes more soluble in the solvent.

For the comparison of liquid phases with different molar masses, apparent partition coefficients are expressed in Pa m^3 (absorbent)/mol (compound absorbed).

$$H = \frac{y \times P}{C_{liq}} \quad (4)$$

Measurement uncertainties were quantified on the basis of the uncertainties of the measuring equipment. Uncertainty propagation was calculated as follows (Eq. (5)):

$$\Delta H = H \sqrt{\left(\frac{\Delta P_{s,eq}}{P_{s,eq}}\right)^2 + \left(\frac{\Delta y_{i,eq}}{y_{i,eq}}\right)^2 + \left(\frac{\Delta V_{liq}}{V_{liq}}\right)^2 + \left(\frac{\Delta n_{i,trans}}{n_{i,trans}}\right)^2} \quad (5)$$

Selectivity, defined as the ratio of the apparent Henry's law constants of two gases, is used to assess the efficiency of absorption of one gas relative to another. At lower selectivities, the first gas is more efficiently absorbed than the second. In this study, the absorption efficiency for CH_4 was compared with that for CO_2 (Eq. (6)).

$$S = \frac{H(\text{CH}_4)}{H(\text{CO}_2)} \quad (6)$$

3 Results and discussion

3.1 Functionalization of phytosterols by carbonation chemistry

This pathway involves the synthesis of amphiphilic phytosterols by a ring-opening oligomerization reaction of glycerol carbonate (GC) initiated by the phytosterols. The targeted reaction is the opening of the GC, assisted by the OH function present on phytosterols, as presented in Figure 1. These hypothetical reaction schemes are based on published

data for the oligomerization of GC (Mouloungui *et al.*, 2006). The cyclocarbonate group is sensitive to nucleophilic attacks. Aromatic alcohols tend to attack the alkylene carbon, whereas aliphatic alcohols are more likely to attack the carbonyl carbon, in accordance with the HSAB principle (Pearson's acid-base concept) (Clements, 2003; Yoo *et al.*, 2001). Kinetically, carbonyl attack (Fig. 1, pathway 1) is favored over alkylene attack (Fig. 1, pathway 2). However, this carbonyl attack is reversible and the product obtained is not stable. Conversely, thermodynamically, the cyclic monomer (more stable) is favored over the polymer and the attack on the alkylene carbon atom is irreversible and accompanied by a loss of CO_2 . The most likely mechanism therefore includes both attack on the alkylene carbon, and attack on the carbonyl carbon (Sonnati *et al.*, 2013).

We synthesized PS-amph in a one-pot reactor with ZnSO_4 as the catalyst, resulting in the production of various molecules (Fig. 2).

FTIR analysis of the native phytosterols and the product (PS-amph) showed a qualitative conversion of functional groups. Reaction with GC increased the intensity of the hydroxyl group peak at 3400 cm^{-1} markedly (Figs. 3A and 3C). This increase indicates an addition of $-\text{OH}$ groups from GC to the phytosterol molecule. Surprisingly, the PS-amph spectrum does not contain the linear carbonate peak at 1730 cm^{-1} predicted on the basis of the theoretical reaction pathway (Fig. 3C and Fig. 2GC). Instead, a strong peak is observed at 1785 cm^{-1} , corresponding to the cyclic carbonate group, with GC eliminated during liquid-liquid extraction. The intensity of this peak can be explained by the formation of coproducts also observed on MALDI-TOF (Fig. 2X and Y). Co-products X and Y, resulting from a transcarbonatation reaction, are problematic as they compete with the intended pathway and inhibit further branching of PS-amph. The rest of the spectrum ($700\text{--}1400 \text{ cm}^{-1}$) corresponds to phytosterol peaks (1464 , 1372 and 1053 cm^{-1}) or GC (1177 and 1082 cm^{-1}).

The two types of phytosterols predominantly present in native phytosterols are campesterol and β -sitosterol, with molar masses of 400 and 414 g/mol respectively (gas chromatography analysis, data not shown). Mass spectrometry of the products formed by the chemical reaction revealed peaks at 400 , 414 and 118 g/mol , indicating the presence of unmodified phytosterols and unreacted GC. Peaks between 473 and 828 g/mol were also observed, corresponding to the expected molecules with n and m between 0 and 5 . The number of n and m branches can be determined from the molar mass indicated by the peak. We found that the maximum values of n and m were close (5 and 3 , respectively), but n was generally

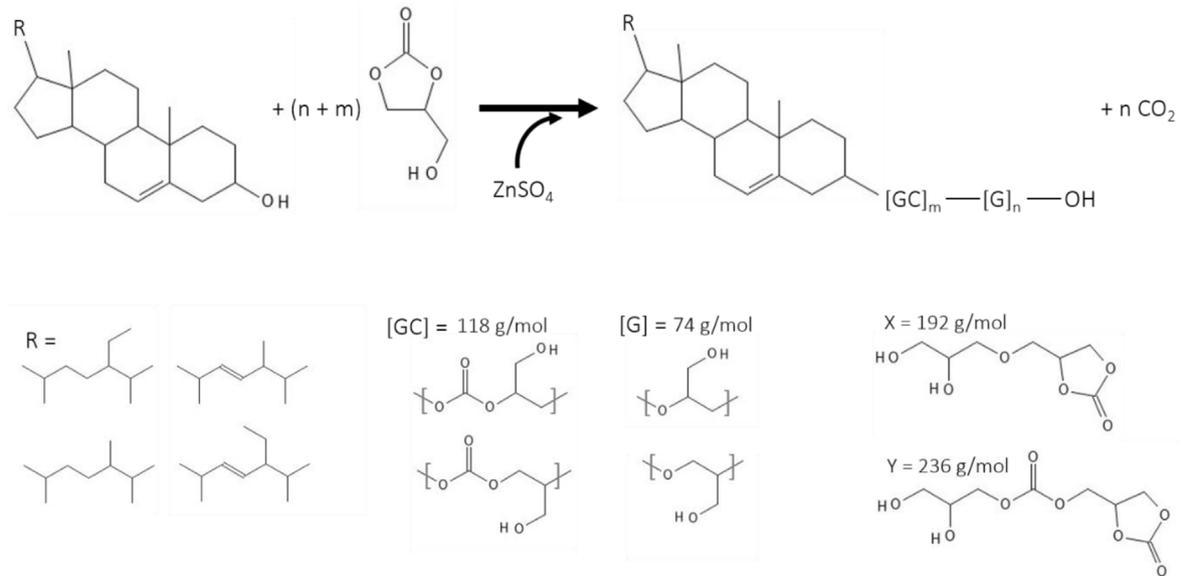


Fig. 2. Chemical pathway and expected molecules.

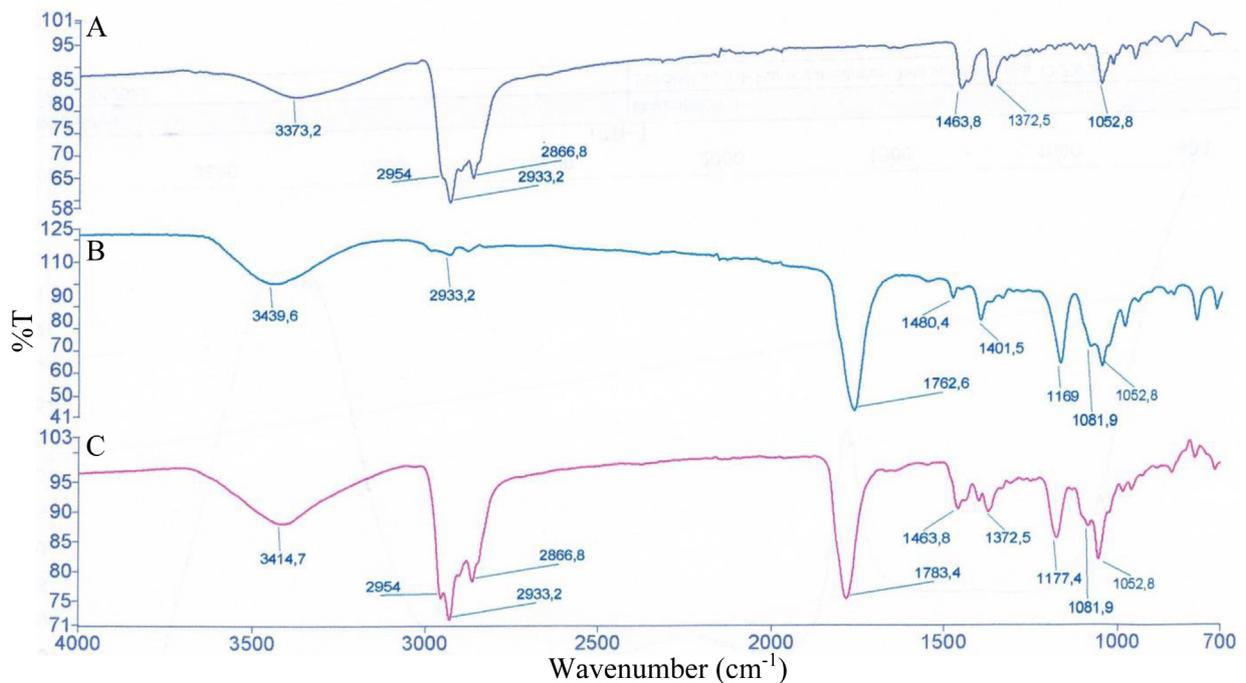


Fig. 3. FTIR spectra of (A) native phytosterols, (B) glycerol carbonate, (C) amphiphilic phytosterols synthesized.

greater than m (Fig. 4). Only a few linear carbonates branched, accounting for their characteristic peak being barely visible on FTIR.

3.2 Characterization of phytosterol dispersions

Dispersions of PS-amph were analyzed by DLS to compare the size distribution of the particles formed by raw mixing (RM)

and by antisolvent precipitation (ASP) (Fig. 5). Granulometry indicated that two populations of particles were present in both cases. Both methods generated a first population with a mean diameter of 2 μm , but RM dispersions then generated a second population of 20 μm -diameter particles whereas ASP dispersions generated a second population of 12 μm -diameter particles. The widths of the peaks indicate that the RM particles were more polydispersed than the ASP particles.

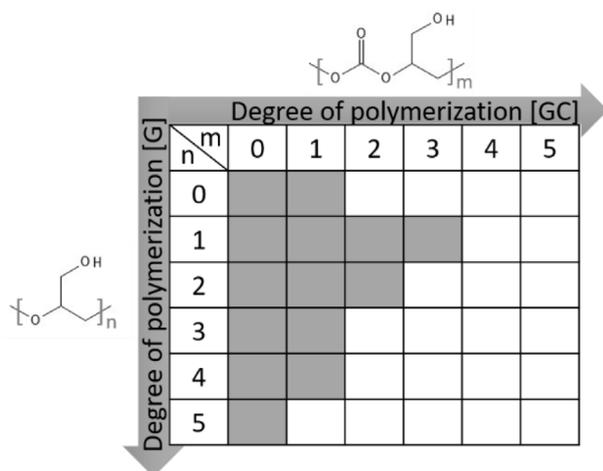


Fig. 4. Degree of polymerization of amphiphilic phytosterols. Number of glycerol [G] and glycerol carbonate [GC] entities was determined by MALDI-TOF analysis.

3.3 Evaluation of the capacity to capture methane and carbon dioxide

The partition coefficient values for CH₄ and CO₂ in several media were compared for the RM dispersions and for the ASP system. Epoxidized soybean oil, commercial olive oil, glycerol carbonate and ethanol were tested in addition to phytosterols dispersions, to compare the different systems.

3.3.1 Raw mixing dispersions (RM)

The affinity of a compound for a given phase can be evaluated by measuring its apparent Henry's law constant *H*. This constant links the partial pressure of the compound in the gas phase to its molar fraction in the liquid phase at liquid-vapor equilibrium. As *H* decreases, the absorption of the compound in the liquid phase increases, reflecting the increase in solubility. CH₄ is known to be poorly soluble in water, which is confirmed by its high *H* value (Fig. 6A). In comparison, CO₂ had a much lower *H* value in water (Fig. 6B). The selectivity defined as the ratio of *H*(CO₂) by *H*(CH₄) was well below 1 (Fig. 6C), which confirms that CO₂ is much more soluble in water than CH₄.

The value of *H* was compared between different systems containing phytosterols in water. For RM systems, *H*(CH₄) decreased to 20 kPa m³ mol⁻¹ as opposed to the 57 kPa m³ mol⁻¹ obtained in water, for both RM-native and RM-amph (Fig. 6A). No significant difference was observed between native and amphiphilic phytosterols. In addition, a solution of 50% water and 50% GC was tested to evaluate the influence of GC on *H*(CH₄). An *H* value of 38 kPa m³ mol⁻¹ was obtained, which is lower than the value obtained for water, but higher than RM-amph. These results indicate that the phytosterol part of the molecule has significantly decreased the *H*(CH₄), thereby improving the affinity of CH₄ for the aqueous phase. However, chemical modification of the phytosterol molecule had no influence on *H*(CH₄). Granulometry showed that amphiphilic phytosterol formed monodisperse particles in

water whereas native phytosterols dispersions polydisperse with presence large aggregates. We expected the amphiphilic phytosterols to have large surface area which could promote interactions with CH₄ resulting in a higher CH₄ solubilization. However, the results showed similar *H*(CH₄) value between RM-native and RM-amph suggesting that the hydrophobic terpenic part of the molecule is responsible of the interaction with CH₄ and that the presence of polyglycerol on the phytosterol structure has no impact on *H*(CH₄).

In comparison with water, *H*(CO₂) slightly increased in systems containing native phytosterols, PS-amph or GC, with no significant difference between the three systems (Fig. 6B). This result suggests that phytosterols do not increase CO₂ solubilization in water. The selectivity *S* of RM-native and RM-amph systems was higher than water (H₂O) and water-GC (50% H₂O 50% GC) (Fig. 6C), which reflects a better CH₄-CO₂ separation due to a higher solubility of CH₄ and no change in the solubility of CO₂.

3.3.2 Antisolvent precipitation system (ASP)

ASP-dispersions were prepared using ethanol at 10 or 50%, which is known to have significant influence on CH₄ solubility (García-Aguilar *et al.*, 2011; Lannung and Gjaldbaek, 1960; Wise *et al.*, 2016). For this study we measured *H*(CH₄) and *H*(CO₂) for 10, 50 and 100% ethanol solutions to evaluate this influence (Figs. 7A and 7B). Results showed that ethanol greatly decreased *H*(CH₄) from 57 kPa m³ mol⁻¹ in pure water to 6.2 kPa m³ mol⁻¹ in pure ethanol. However, *H*(CO₂) was slightly higher for 10 and 50% ethanol, and significantly decreased for 100% ethanol. The resulting selectivity increases when ethanol increases (Fig. 7C).

For systems with 10% ethanol, the presence of 1 g/L particles of native phytosterols (ASP-native 1) had no significant effect on *H*(CH₄) or *H*(CO₂) compared with the 10%-ethanol solution (Figs. 8A and 8B). However, the presence of 1 g/L particles of PS-amph (ASP-amph 1) halved the value of *H*(CH₄) and slightly decreased *H*(CO₂). The addition of amphiphilic phytosterol microparticles significantly improved the selectivity of absorption between CH₄ and CO₂. The nature of the particle seems to have an impact on the interaction with CH₄, as ASP-amph particles are covered with polyglycerol groups whereas ASP-native particles are covered with Tween80.

In order to increase the phytosterol concentration beyond 1 g/L, ethanol concentration also had to be increased due to poor solubility of phytosterols. For this reason, ASP-native 5, ASP-amph 5 and ASP amph 15 were composed of 50% ethanol. Dispersions with PS-amph particles (ASP-amph 5 and 15) had similar constants for CH₄ and CO₂ with values between 27 and 30 kPa m³ mol⁻¹ for *H*(CH₄) and about 2.8 kPa m³ mol⁻¹ for *H*(CO₂) (Figs. 9A and 9B). No significant difference was observed when increasing the particle concentration by 3-fold, and more importantly no significant difference was observed with the control. The presence of particles with amphiphilic phytosterols had no significant impact on CH₄ and CO₂ solubility compared with the presence of 50% ethanol alone.

For dispersions with native phytosterols (ASP-native 5), the values of *H*(CH₄) and *H*(CO₂), 12.7 kPa m³ mol⁻¹ and

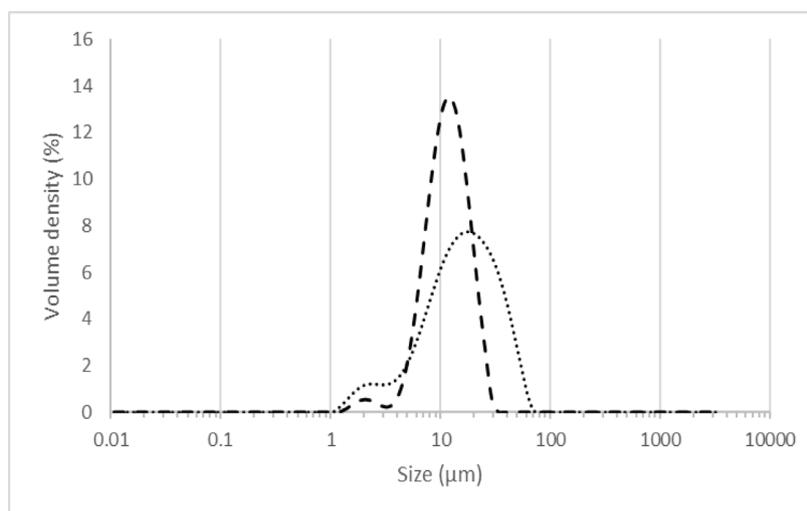


Fig. 5. Size distribution of particles generated by the raw-mixing (RM) method (●●●) or by antisolvent precipitation (ASP) (---) with 1 g/L amphiphilic phytosterols.

$1.2 \text{ kPa m}^3 \text{ mol}^{-1}$, were half those obtained for the corresponding control (Figs. 9A and 9B). The presence of Tween 80 in ASP-native samples may form micelles of surfactant, providing polar and nonpolar environments that increased CH_4 solubilization (García-Aguilar *et al.*, 2011). At low surfactant concentration (ASP-native 1), this phenomenon was not observed, but for high concentration (ASP-native 5) CH_4 solubility was significantly reduced.

Overall, our observations suggest that the presence of phytosterols increased the affinity of methane for aqueous solutions under certain conditions. One possible reason for this is that methane is adsorbed onto the surface of the particles. On increasing particle concentration, the surface area for adsorption increased as well. We therefore expected to solubilize more CH_4 illustrated by a lower $H(\text{CH}_4)$. However, for PS-amph dispersions, both $H(\text{CH}_4)$ and $H(\text{CO}_2)$ remained constant even when particle concentration was multiplied by three. For native phytosterols dispersions, $H(\text{CH}_4)$ decreased strongly between concentrations of 1 and 5 g/L, but this decrease probably reflects the presence of ethanol and surfactant. Because of the influence of ethanol on CH_4 solubility, we cannot evaluate on the impact of higher concentrations of phytosterol particles on CH_4 solubilization. More work is necessary to remove ethanol without degrading the particles to study the absorption capacity of these systems with pure particles.

According to the same hypothesis, we also expected that increasing the hydrophilicity of phytosterols would result in a greater surface area available for interactions with CH_4 , resulting in a decrease in $H(\text{CH}_4)$ PS-amph. For systems with 10% ethanol, better CH_4 solubility was indeed observed when using amphiphilic phytosterols relative to native phytosterols. Again, in systems containing 50% ethanol, no such improvement was observed probably because of the strong ethanol influence on CH_4 solubility.

Although phytosterols influenced methane solubility in water, $H(\text{CH}_4)$ could not be decreased beyond $20 \text{ kPa m}^3 \text{ mol}^{-1}$, unless using other components such as ethanol or Tween80. Phytosterols showed to have specific interaction with CH_4

and not with CO_2 but systems containing phytosterols had a selectivity for methane of 0.16 at best, where a selectivity greater than 1 would be more convenient for efficient CO_2 - CH_4 separation.

Increasing the concentration and modification of the surface of the particles did not have significant impact on methane solubility. These findings call into question the hypothesis that particles adsorb methane on their surface and suggest that the improvement in the affinity of methane for aqueous systems probably results from another physical phenomenon that could involve solubilization and diffusion.

3.3.3 Raw mixing with oily systems

Finally, we studied the affinity of methane for absorbing phases containing larger amounts of fat, such as a phase containing 20% epoxidized soy oil and another phase consisting of pure olive oil (Fig. 10). Both oils have been reported to contain phytosterols (Yang *et al.*, 2019). The liquid phase containing 20% soy oil decreased $H(\text{CH}_4)$ to $19.7 \text{ kPa m}^3 \text{ mol}^{-1}$, a similar value compared with RM-native and RM-amph which contained only 1.6% of phytosterols. Pure olive oil was the only absorbing phase that reduced $H(\text{CH}_4)$ below $10 \text{ kPa m}^3 \text{ mol}^{-1}$. Both oils slightly increased CO_2 solubility, but selectivity obtained using olive oil was the highest of all the solution tested in this study.

4 Conclusion

We used microparticles of phytosterol to improve the solubility of methane in aqueous solution. Amphiphilic phytosterols were synthesized with glycerol carbonate to render them more hydrophilic. Mass spectrometry analyses showed that up to five branches were added to the molecule, and FTIR showed an increase in the number of hydroxyl groups. The objective of this study was to identify a solution to improve CH_4 absorption, and to increase the selectivity of absorption between CH_4 and CO_2 . Different dispersions containing phytosterols, dispersed by raw mixing or by antisolvent precipitation, were tested. The presence of these

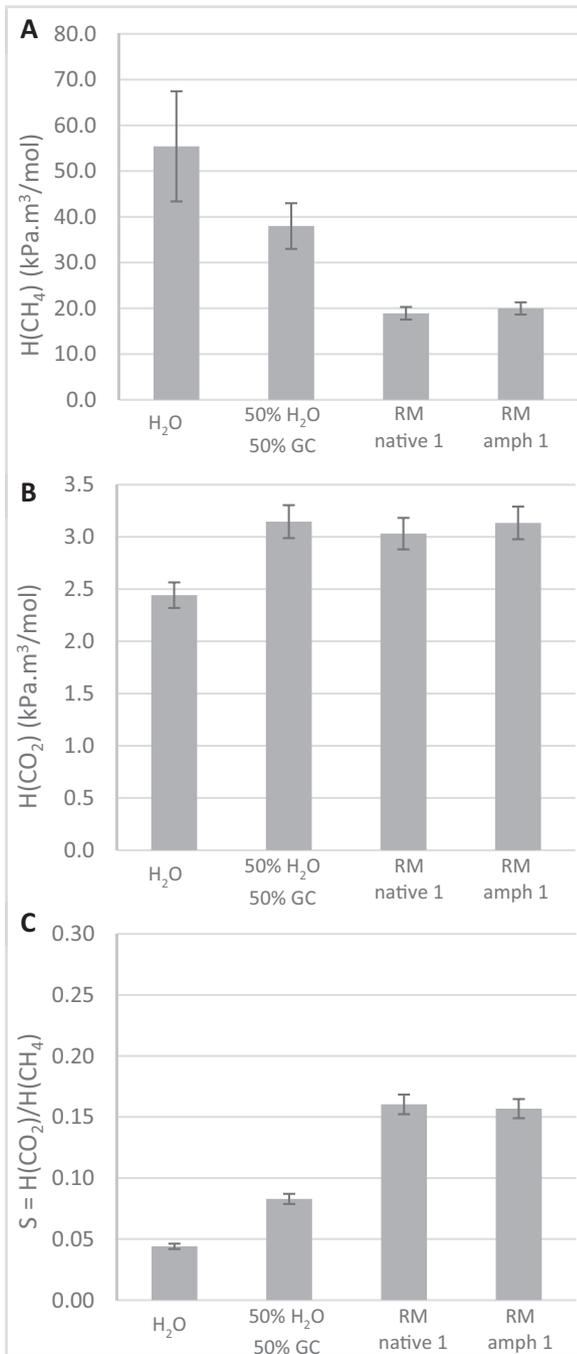


Fig. 6. Methane (A) and carbon dioxide (B) absorption and selectivity (C) results for dispersions generated by raw mixing from 1.6% native or amphiphilic phytosterols for RM-native and RM-amph, respectively.

particles of 2–20 μm in diameter in aqueous solutions decreased the apparent partition coefficient of CH₄ between gas and liquid. RM dispersions decreased H(CH₄) without modifying H(CO₂), resulting in a four-fold improvement in selectivity for CH₄. ASP dispersions also had a lower H(CH₄) and better selectivity if amphiphilic phytosterols were used. We could not evaluate the effect of the phytosterol concentration because of the strong influence of ethanol on CH₄ solubilization. Surprisingly, chemical modification of the

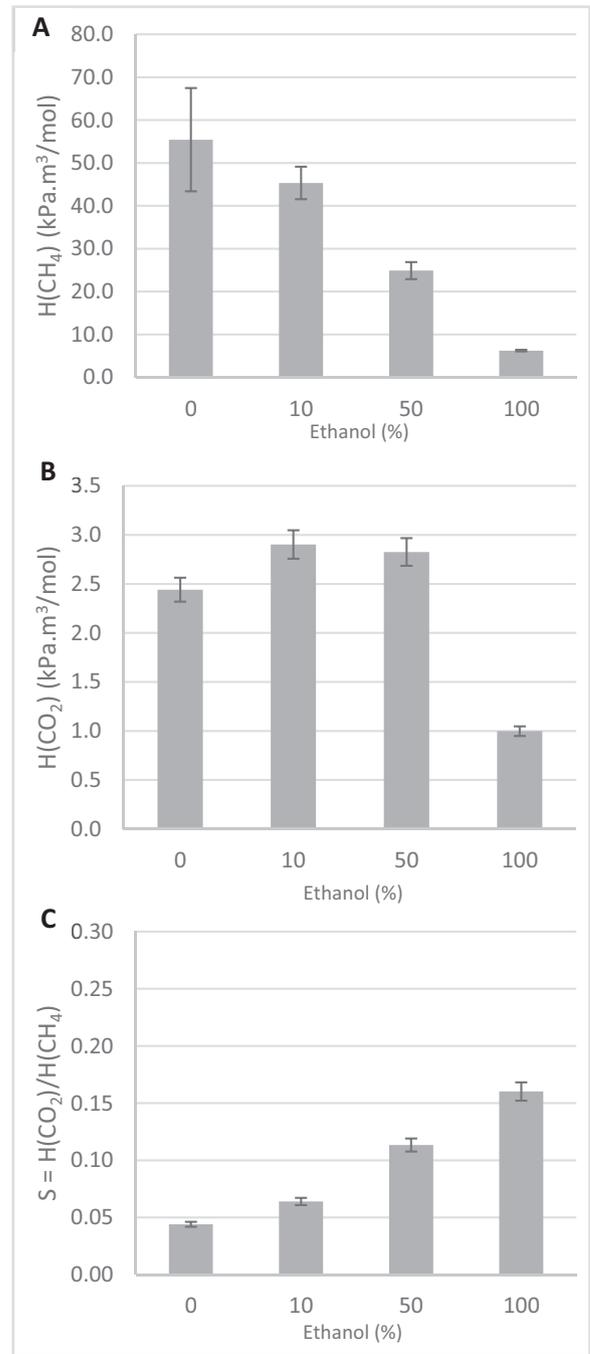


Fig. 7. Methane (A) and carbon dioxide (B) absorption and selectivity (C) results for solutions containing different proportions of ethanol in water.

phytosterol had no significant effect on the improvement in methane solubility when the raw-mixing method was used. When monodisperse particles were formed by antisolvent precipitation, CH₄ showed a better affinity with systems containing amphiphilic phytosterols than native phytosterols and Tween 80. Pure olive oil had the best capacity for absorption, with the lowest H(CH₄) of all absorbing phases tested in this study. Overall, these results suggest that phytosterols, crude biosourced compounds, are of great interest for CH₄ solubilization. The use of phytosterols for

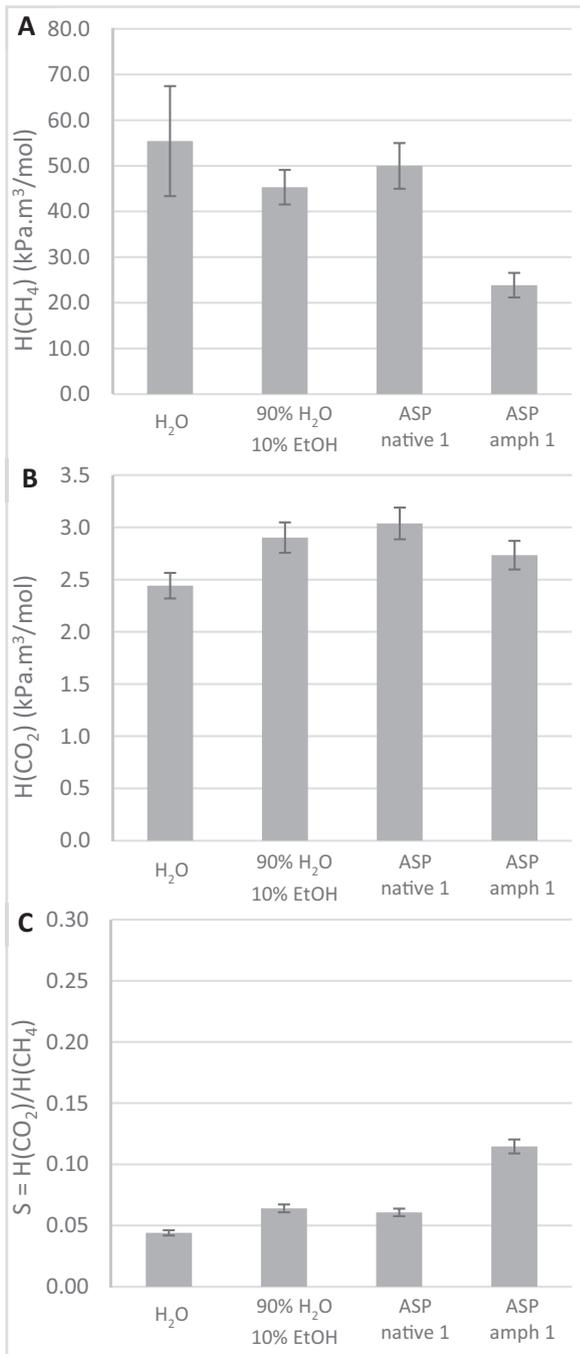


Fig. 8. Methane (A) and carbon dioxide (B) absorption and selectivity (C) results of solution containing particles and 10% ethanol, comparison with water.

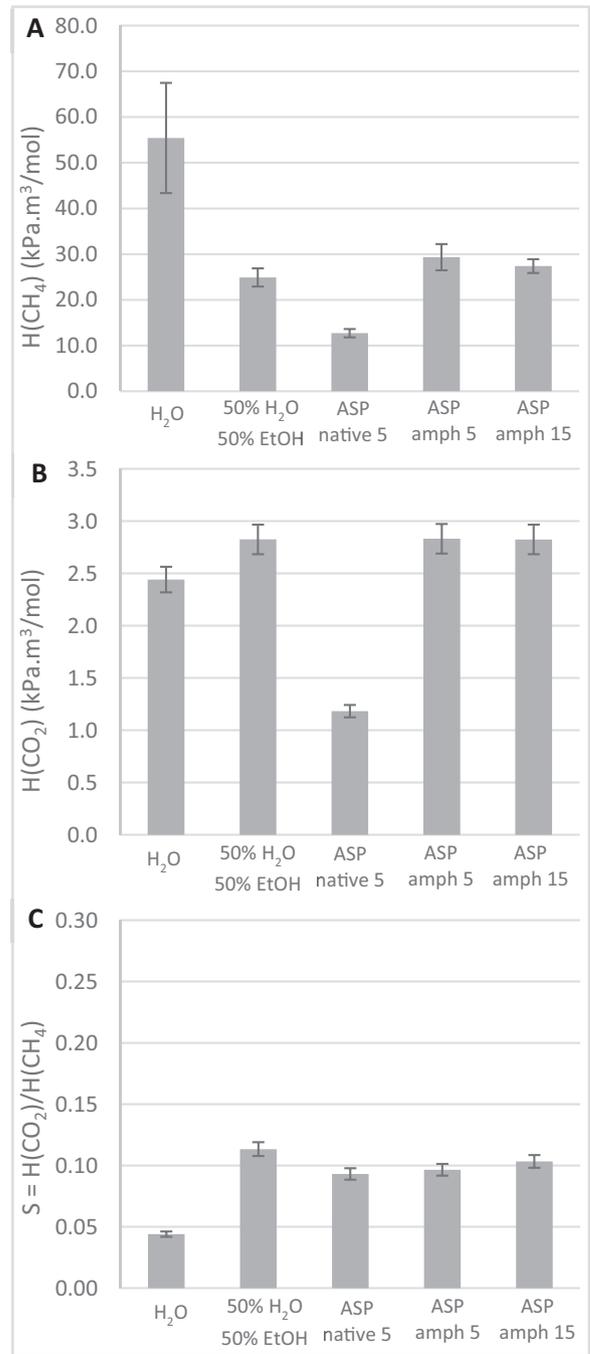


Fig. 9. Methane (A) and carbon dioxide (C) absorption and selectivity (C) results of solution containing particles and 50% ethanol, comparison with water.

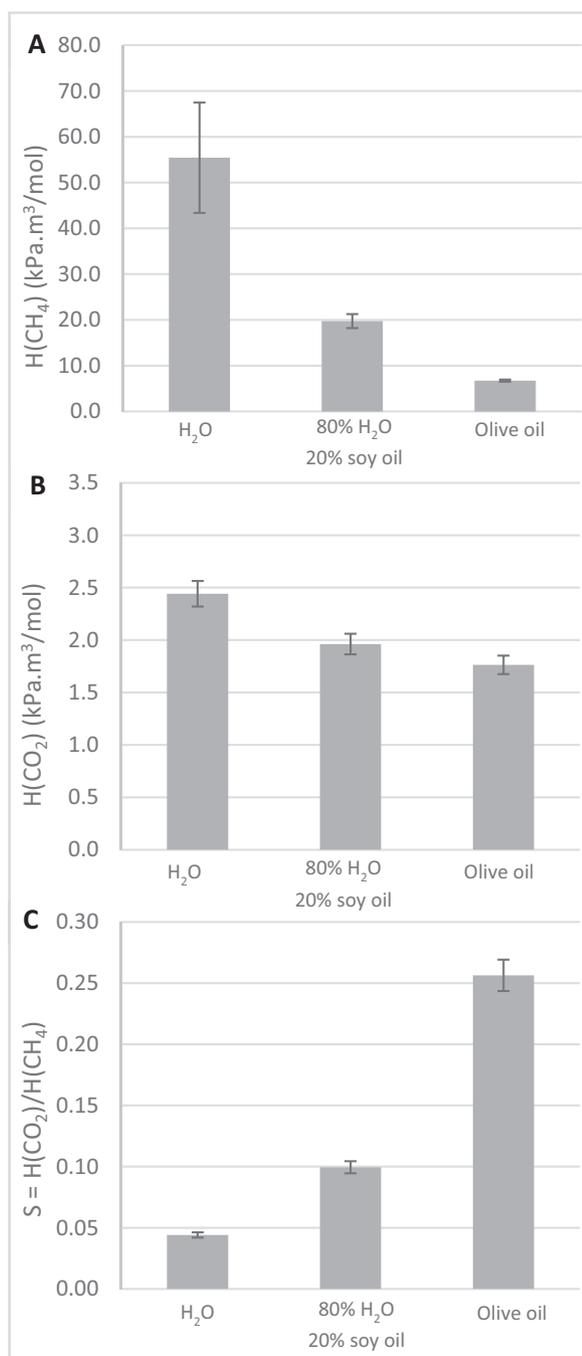


Fig. 10. Methane (A) and carbon dioxide (B) absorption and selectivity (C) with oils as the absorbing phase.

this purpose was never reported in the literature before, which makes this approach innovative and useful. Phytosterol particles as additives could help improve existing methods for CH₄ capture and CH₄-CO₂ separation processes instead of petrochemical surfactants.

Conflicts of interest

The authors declare that they have no conflicts of interest in relation to this article.

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